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### The Evolution of Pollution Resistance in the Atlantic Killifish

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### Rapid Evolution of Pollution Resistance in Atlantic Killifish Matthew Rock<sup>1</sup>, Jeffrey Markert<sup>1</sup>, Bryan Clark<sup>2</sup>, Diane Nacci<sup>2</sup> <sup>1</sup> Department of Biological Sciences – Providence College – Providence, Rhode Island (USA) <sup>2</sup> U.S. Environmental Protection Agency – ACESD – Narragansett, Rhode Island

## Summary

We examined genetic variation in *Fundulus heteroclitus* populations to better understand their ability to persist in Superfund sites like New Bedford Harbor (NBH) (MA, USA) which is contaminated with a mix of organic toxicants and heavy metals. We isolated DNA from killifish along the Rhode Island and Massachusetts coast at 11 different locations. We identified allele frequency differences at loci consistent with this adaptation. Overall, we observed the expected pattern of genetic isolation by distance when all loci were examined as a group. However, genotypes at some loci are consistent with selection at polluted locations. As predicted by other researchers, loci associated with the Aryl Hydrocarbon Receptor (AHR) system show large differences in allele frequencies between polluted and non-polluted sites.

## Variation and Clustering



# Background

New Bedford Harbor (NBH) is contaminated with a mix of organic toxicants including Polychlorinated Biphenyls and heavy metals released in the early 20<sup>th</sup> century. PCBs harm many tissues, including developing hearts, causing 'tube heart'. However, a large population of Atlantic Killifish thrives in New Bedford Harbor. Lab experiments show that this tolerance is heritable (Nacci et al, 2010).



~500 DNA samples from 11 sites were genotyped at 192 putative SNP loci using a Fluidigm EP-1 system. Of these, 55

# Amino Acid Changes

*Fundulus heteroclitus* embryonic development: 0 normal embryo, 1 & 2 PCB exposed embryos (Di Giulio and

Minor alleles at 27 of the 55 SNPs are consistent with missense or nonsense mutations, not all loci code for known proteins. Many of these are part of the AHR pathway.

NP_NAME	Common Amino Acid	Common Condon	Alternate Amino Acid	Alternate Codon	Mutation Type
HR1_161	Alanine	GCC	Glycine	GGC	Missense
HR1_1530	Glycine	GGG	Arginine	CGG	Missense
HR1_2289	Methionine	ATG	Isolucine	ATC	Nonesense
HR2b358573	Glutamic Acid	GAG	Lysine	AAG	Missense
and the second					

Sampling locations with sediment PCB concentrations (Nacci et al, 2005) along the coast of Rhode Island and Massachusetts



STRUCTURE analysis (above) suggests 3 genetically distinct units spread across the 11 collection sites. The red cluster is associated with higher levels of sediment PCBs at NBH, and perhaps evolved from the eastern cluster in the mid 20<sup>th</sup> century

**Principal Components Analysis with Adegenet/DAPC** The first Principal Component (X axis) explains 43% of the overall variation and is consistent with geographic variation. The second Principal Com-ponent explains 21% of the variation. Allele loadings on this second axis cause us to hypothesize that this variation reflects population genetic changes in response to PCBs in the sediment.





G<sub>ST</sub>

AHR1\_161 AHR1\_948 AHR1\_1530 AHR1\_2289 AHR2\_792 AHR2\_792 AHR2\_792 AHR2\_1929 AHR25368573 ARNT\_518094 CYP1A\_2140 CYP1A\_2140 CYP3A\_1166 FABP Hepcidin2\_399 LDH8654 LDH81033 NADH10\_107 NADH10\_107 NADH10\_109 Thioredoxin\_582(ight)





Changes in the amin<sup>b</sup> acid sequence of the protein coding regions leading to missense and nonsense mutations.

CYP1A

CYP1B1

GST1a

NQ01

AHRR





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developed data analysis tools.